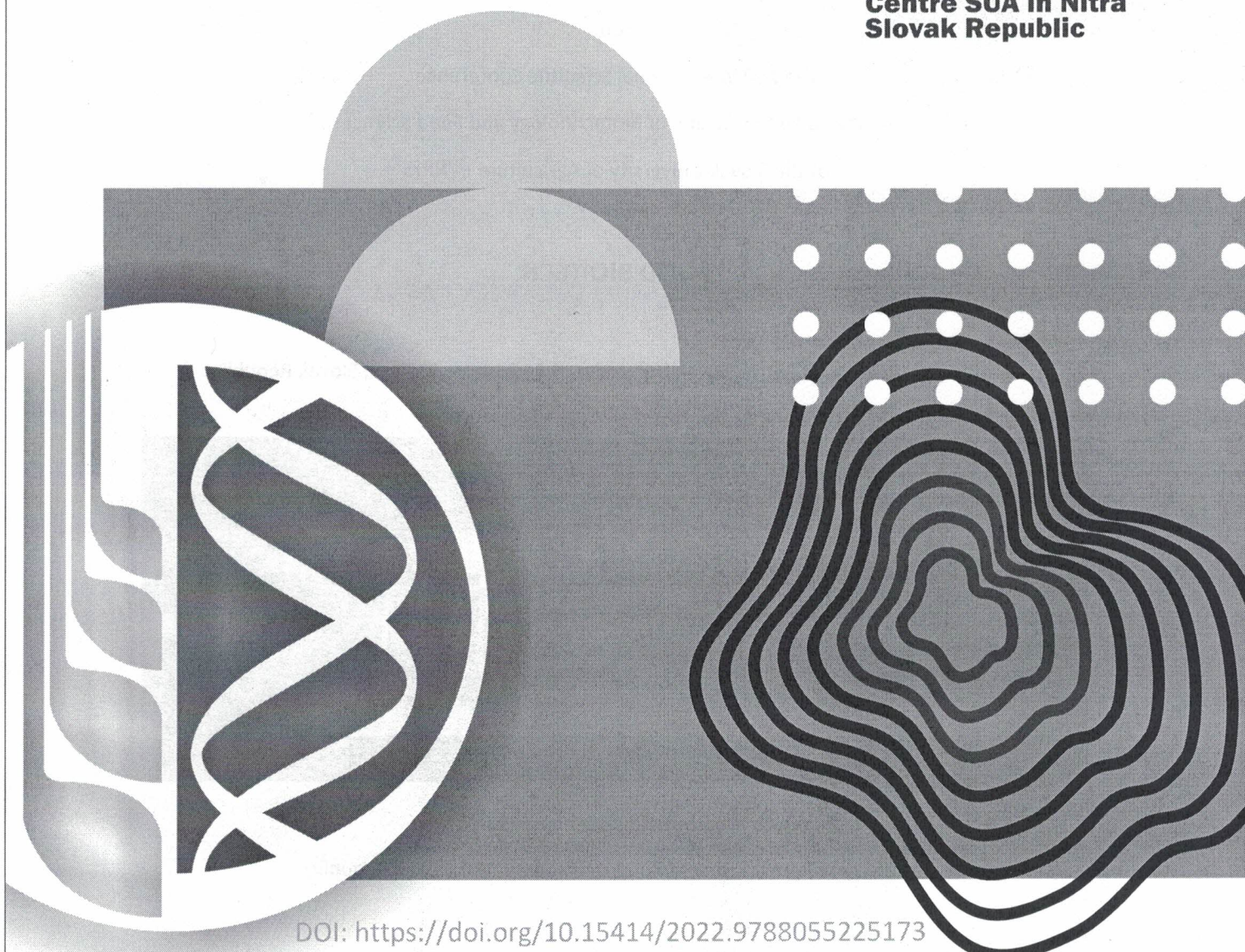


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DOI: <https://doi.org/10.15414/2022.9788055225173>

BOOK OF ABSTRACTS

**FACULTY OF BIOTECHNOLOGY
AND FOOD SCIENCES**

SLOVAK UNIVERSITY OF AGRICULTURE IN NITRA

FOOD | BIO | TECH 2022

5th - 6th October

Slovak University of Agriculture in Nitra
Nitra, SLOVAK REPUBLIC

BOOK OF REVIEWED ABSTRACTS

from

the 15th International Scientific Conference

organized by the Faculty of Biotechnology and Food Sciences

of the Slovak University of Agriculture in Nitra

FOOD | BIO | TECH

AgroBioTech Research Centre, Slovak University of Agriculture in Nitra, Slovak Republic,
October 5th – 6th 2022



which is being held under auspices of the
Ministry of Agriculture and Rural Development of Slovak Republic

Nitra, 2022

DOI: <https://doi.org/10.15414/2022.9788055225173>

Title: Book of Reviewed Abstracts from the 15th International Scientific Conference Food|Bio|Tech,
Faculty of Biotechnology and Food Sciences, Slovak University of Agriculture in Nitra, Slovak Republic

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**THIOREDOXIN FUSION TAG IMPROVES PROTEIN YIELD OF RECOMBINANT
DROSERA BINATA GLUCANASE****Miroslav Rajnivec, Jana Libantová**

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The production of soluble recombinant proteins in *Escherichia coli* remains one of the major bottlenecks of prokaryotic expression systems. The addition of fusion tags to enhance the solubility of the produced protein is one of the best solutions for expressing enzymatically active eukaryotic proteins. In our work, we over-expressed the basic β -1,3-glucanase from the carnivorous plant *Drosera binata* in *Escherichia coli* cells. Two forms of enzymatically active protein were produced, which differ in the presence (DbGluc+Trx, ~50 kDa) or absence (DbGluc-Trx, ~30 kDa) of the thioredoxin fusion tag. Both forms of β -1,3-glucanase also contain the 6xHis-tag sequence for purification by affinity chromatography. Successful production of both forms of recombinant β -1,3-glucanase was detected by SDS-PAGE and the presence of both forms of β -1,3-glucanase was confirmed by on-gel detection of Ni-NTA conjugated fluorescence dye signal. Although both forms of β -1,3-glucanase showed similar specific activity ($68.12 \text{ U}\cdot\text{mg}^{-1}$ DbGluc+Trx and $64.61 \text{ U}\cdot\text{mg}^{-1}$ DbGluc-Trx), thioredoxin fusion tag increased protein yield of β -1,3-glucanase two-times during purification on Ni-NTA agarose. We estimated that 69% of total enzymatic activity was preserved after purification of DbGluc+Trx, in comparison with DbGluc-Trx, where β -1,3-glucanase retained only 31% of its activity. Thioredoxin fusion tag significantly increased the yield of recombinant β -1,3-glucanase and its presence during purification steps is crucial for sufficient transgene production.

Keywords: *Drosera binata* β -1,3-glucanase, Ni-NTA purification, thioredoxin fusion tag

Acknowledgements: This work was co-funded by a grant from the Slovak Grant Agency VEGA 2/0041/20 and Cost Action CA18111—Genome editing in plants—a technology with transformative potential. This publication was also supported by the Operational program Integrated Infrastructure within the project Demand-driven research for sustainable and innovative food, Drive4SIFood 313011V336, co-financed by the European Regional Development Fund.